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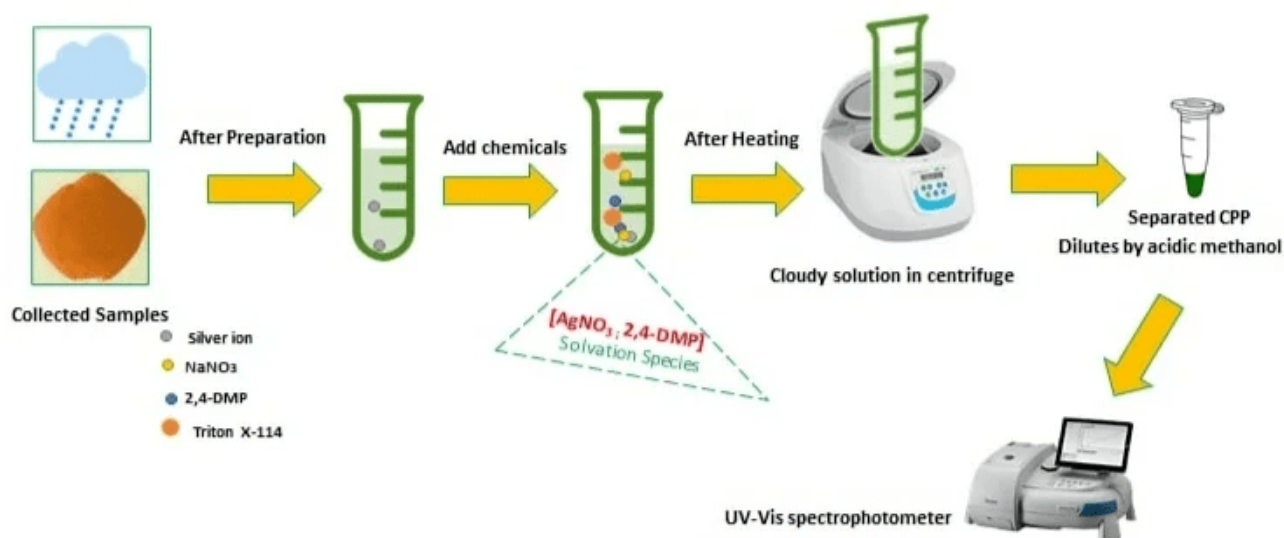
Methodology for Preconcentration and Determination of Silver in Aqueous Samples using Cloud Point Extraction

Ebaa Adnan Azooz¹ , Ghusoon Jawad Shabaa² , Emad Abbas Jaffar Al-Mulla^{*3}  

¹Department of Chemistry, The Gifted Students' School in Najaf, Ministry of Education, Iraq

²College of Medicine, Jabir Ibn Hayyan Medical University, Najaf, Iraq

³Pathological Analysis Techniques, Faculty of Health and Medical Techniques, Al-Furat Al-Awsat Technical University, Kufa, An-Najaf, Iraq



For the selective extraction of silver, a cloud point extraction (CPE) procedure was developed. After synthesizing the solvation species through the reaction of silver ions with 2, 4-dimethyl pentane-3-one (2,4 DMP), the salting-out agent ($0.4 \text{ mol L}^{-1} \text{ NaNO}_3$) was added at $35 \text{ }^\circ\text{C}$ and, after 10 min, Triton X-114 was used to separate silver ions from aqueous solution. The type and quantity of salting-out agent, silver ion, temperature, heating time, and surfactant volume were all examined as important factors determining the CPE. The analytical curve in the $0.1\text{-}100 \text{ } \mu\text{g L}^{-1} \text{ Ag}$ range was straight at optimal conditions. The detection limit (LOD), quantification limit (LOQ), and enrichment factor (E) were $0.05 \text{ } \mu\text{g L}^{-1}$, $0.15 \text{ } \mu\text{g L}^{-1}$, and 200, respectively. The relative standard deviation (RSD) was estimated as 0.2-3.9% ($n = 5$) in relation to 1, 40,

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and $80 \mu\text{g L}^{-1}$ Ag. Flame atomic absorption spectrometry and spectrophotometry exploiting dithizone were used to assess the CPE accuracy. The proposed approach was then applied to river water, rain water and sand samples.

Keywords: cloud point extraction, salting out effect, silver, solvation system, solvent extraction.

INTRODUCTION

Non-polluting and accurate methods for trace element determination are desirable [1,2]. To remove and extract tiny quantities of elements, a variety of approaches have been utilized [3-5], including solid-phase extraction [6,7], liquid-phase extraction [8-12], and cloud point extraction (CPE) [13-16]. Researchers are interested in CPE because it follows green chemistry principles. After all, the chemicals utilized are tiny and non-toxic. CPE also low-cost, accurate, efficient, and quick [17-22]. Achieving a critical micellar concentration (CMC) requires a cloud point temperature that is just right. Cloud point phase (CPP) (non-hydrophilic, with large micelle quantity and tiny volume) and an aqueous phase (large volume, hydrophilic nature) are produced as a result. To create hydrophobic complexes, target components are combined with an organic chelating reagent, which is further transported towards CPP [23-25]. Solvation molecules have hydrophobic properties and can be equally distributed in the micelle phase [13]. In the condition of salting-out processes, nitrate and hydrophobic reagents [$\text{M}(\text{NO}_3)_x$; b (organic reagent)] are combined. Many agents, including aldehydes, ketones, esters, etc. Generally, the salting-out process is caused by ions desorbing from the hydrophilic portions of micelles, enhancing inter-micelle attraction and, as a result, causing surfactant molecules to precipitate. The salting-out effect was absent and presence of NaNO_3 , KNO_3 , and other chemicals. The solution ionic strength is mostly influenced by nitrate salts, which reduce the hydration shell surrounding metallic ions and aid in the transport of ionic species to CPP [26-28].

Silver is antibiotic, thermally and electrically conductive. Silver compounds are commonly utilized in water purification filters, medical materials, food, and advanced materials. Because the amount of silver in the crustal ground is low, water naturally includes trace amounts of it. As a result, a particularly sensitive methodology should be utilized for silver detection in water for evaluating the toxicity effect in bio-organisms [30,31]. In this instance, an ultrasound-assisted cloud point extraction procedure was implemented. Silver was extracted from samples using copolymer and Fe_3O_4 nanoparticles at pH 5.0, and spectrophotometrically detected. The detection limit (LOD) was $0.85 \mu\text{g L}^{-1}$ [32].

The primary goal of this study was to extract and determine silver in water and sand samples using CPE. The surfactant, chelating, and salting-out reagents were Triton X-114, 2, 4-DMP and NaNO_3 , respectively. The factors that influence CPE were examined.

MATERIALS AND METHODS

Reagents and solutions

All the substances (with purity ranging from 99.0 to 99.2%) used in this study were from Merck, Darmstadt, Germany. Deionized water was used in the preparation of all samples and solutions. A standard solution 1.00 g L^{-1} Ag was prepared by dissolving AgNO_3 with water in the presence of 1 mL of HNO_3 . Then, the solution was transferred to a suitable volumetric flask and the volume was completed to the mark using water. The 2,4-DMP solution ($1 \times 10^{-2} \text{ mol L}^{-1}$) was prepared in water which contains drops of surfactant. The NaNO_3 solution (1.00 mol L^{-1}), was prepared in water. The dithizone solution ($1 \times 10^{-2} \text{ mol L}^{-1}$) was prepared in CCl_4 .

Instrumentation

A water bath model Cambridge, England, (WNB7-45), ($\pm 0.0001 \text{ g}$ precision) A and D company, Japan, a UV-Vis Biochrom spectrophotometer Libra, S60 (Cambridge, England), and an AA 6800 Shimadzu flame atomic absorption spectrophotometer (Kyoto, Japan) with a silver hollow cathode lamp were employed.

Test methodology

An aliquot comprising 25 mL of an aqueous solution, $100 \mu\text{g L}^{-1}$ silver ions, 0.4 mol L^{-1} NaNO_3 , $1 \times 10^{-4} \text{ mol L}^{-1}$ 2,4-DMP solution, and 0.5 mL of Triton X-114 were mixed and deionized water was diluted to the line of the 50 mL centrifuge tube. This was done in a water bath to accelerate the formation of the micelle at $35 \text{ }^\circ\text{C}$ for 10 min. The mixture was centrifuged for 12 minutes at 4000 rpm and then refrigerated for 4.0 minutes in the freezer. Then, the hydrophobic phase was separated and 5 mL of an acidic methanol solution containing nitric acid (1.0 mol L^{-1}) was added. Afterward, the absorbance was measured at $\lambda_{\text{max}} = 325 \text{ nm}$. A blank solution was prepared and analyzed in the same way.

To extract silver ions from aqueous solutions, the dithizone (H_2Dz) methodology [41] was employed. The 0.001% H_2Dz in CCl_4 was added to a silver solution containing HNO_3 until the concentration reached $1\text{-}2 \text{ mol L}^{-1}$, then shaken for 5 minutes. The step was fivefold repeated. The color of an aqueous solution after the last amount of H_2Dz has been added should not change. Separation of the silver-containing aqueous layer by lowering the pH to 4-5 with ammonia, then removing the silver in CCl_4 with 0.001% H_2Dz . To exclude any residual H_2Dz from the sample, a dilute ammonia solution (2 drops of conc. NH_3 solution in 25 mL of water) was utilized [42]. The analytical curve was used to determine the amount of silver ions. Equation 1 gives the distribution ratio (D). Log D data is placed in all of the graphics to provide more structured and wide data [33].

$$D = \frac{[\text{Ag(I)}]_{\text{CPP}}}{[\text{Ag(I)}]_{\text{aq}}} \quad (1)$$

Sample Preparation

Water samples

All water samples were filtered through a $0.45 \mu\text{m}$ Millipore filter and stored in a polyethylene container for future application after being acidified to pH 2.0 with HNO_3 to reduce silver ions adsorption on the container surface.

Sand samples

The sand samples were gathered from several sites throughout the Najaf desert in Iraq. The samples were dehydrated and mashed at $90 \text{ }^\circ\text{C}$ for 15 hours before being put through a 130 grit filter. In a Teflon bottle, 5 mL of each HNO_3 , H_2O_2 , and H_2O of the measuring sand were added to 1.5 g and destroyed using a microwave digestion oven with an application cooking program [25]. The extra H_2O_2 was then neutralized using 20 mL of H_3BO_3 (5% w/v). After that, the mixture was filtered, and the volume was filled up to 50 mL. Masking reagents (KCN and $\text{K}_2\text{S}_2\text{O}_3$) were applied to samples at a concentration of 0.1 mol L^{-1} [25].

RESULTS AND DISCUSSION

The spectrophotometric investigation involved 50 mL of aqueous solutions containing $100 \mu\text{g L}^{-1}$ Ag(I) , 0.4 mol L^{-1} NaNO_3 , 1.0% Triton X-114 and $1 \times 10^{-4} \text{ mol L}^{-1}$ 2,4-DMP. The heating temperature was maintained as $35 \text{ }^\circ\text{C}$ for 10 minutes. The extracted components were spectrophotometrically evaluated ($\lambda_{\text{max}} = 325 \text{ nm}$). Effects of the main involved factors were investigated for dimensioning the proposed procedure.

Influence of the sodium nitrate concentration

Aqueous solutions (50 mL) including $100 \mu\text{g L}^{-1}$ silver ions were handled with different quantities of salting-out (NaNO_3) in the range of ($0.1\text{-}1 \text{ mol L}^{-1}$) according to the test methodology at $40 \text{ }^\circ\text{C}$ for 15 minutes. After treating the Ag(I) aqueous solutions as above described, it was realized that 0.4 mol L^{-1} was the most effective concentration of the salting-out agent (NaNO_3). This concentration corresponded to the fastest formation equilibrium with the development of solvation species. This was not attained with a concentration just under or higher than the optimal value. Equation 2 depicts the structure of solvation species. Figures 1 and 2 show the influence of NaNO_3 concentrations on salting-out.

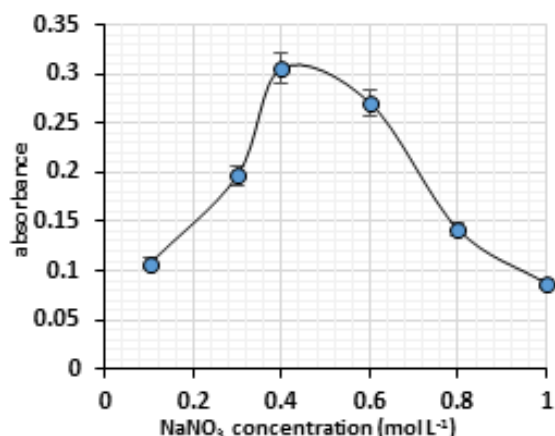


Figure 1. Influence of the NaNO₃ concentration on the extraction efficiency.

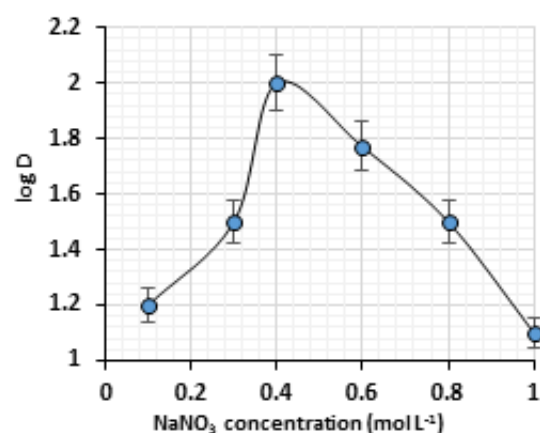


Figure 2. Influence of the NaNO₃ concentration on the D.

Influence of the concentration of the silver ions

Figures 3 and 4 display the effectiveness of silver ion solutions at different concentrations (0.1-100 $\mu\text{g L}^{-1}$) including 0.4 mol L⁻¹ NaNO₃, and completing the task according to the applied methodology. The findings show that silver ions act a parameter that influences the thermodynamic equilibrium of the synthesis solvation species, as well as the transport to the micelle. The optimal concentration amount of silver was then found as 100 $\mu\text{g L}^{-1}$ and this result is in agreement with references [33,41].

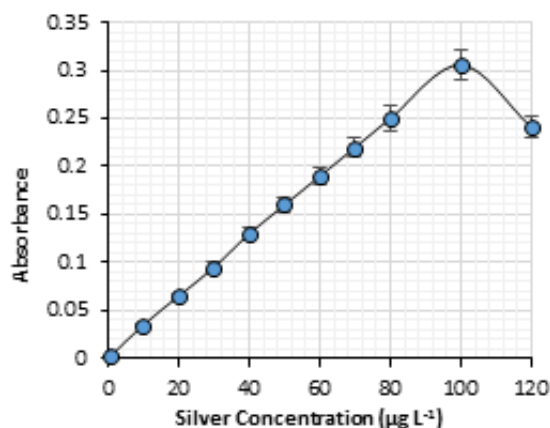


Figure 3. Influence of the silver ion concentration on the absorption of solvation species.

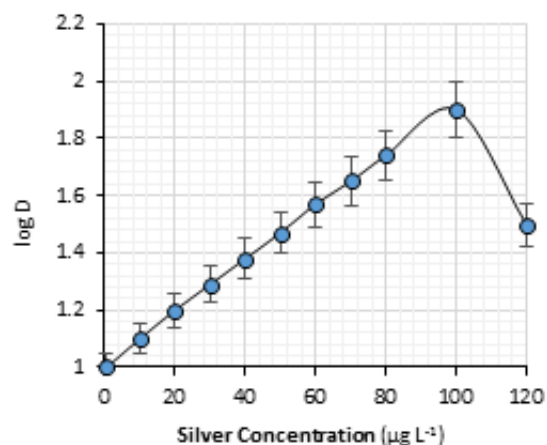


Figure 4. Influence of the silver ion concentration on extraction efficiency.

Influence of the ketone nature

Five different ketones were tested as organic agents (1×10^{-4} mol L⁻¹) for the formation of solvation species with silver nitrate separation according to the test methodology. The results in Table I show a different extraction efficiency using different ketones. The 2,4-DMP appears to be the best value for D and absorbance depends on chemical structure [34]. It increases the lipophilic properties of solvation species, which allows transfer to the CCP for easy separation. It permits the preconcentration at a specific

temperature with the presence of a salting-out impact. 2,4-DMP was then selected to be used in the following studies.

Table I. Effects of various ketones on the extraction efficiency and distribution ratio (D)

Ketones	λ_{\max} nm	Absorbance	D
2,4-DMP	325	0.342	95.460
Acetone	256	0.105	43.461
Acetophenone	291	0.201	54.621
Methyl Iso Butyl Ketone	292	0.247	80.752
Propanone	297	0.233	75.500

Influence of the temperature and heating time

In CPE, temperature and heating time are important parameters. Figure 5 refers to a thorough process for extracting silver ions at various temperatures. The ideal temperature was selected as 35 °C. Thus, it was suitable for forming a hydrophobic CPP and provided enhanced extraction efficiency for silver ion removal [33,34]. Enthalpy, entropy and Gibbs free energy were then determined as $\Delta H_{\text{ex}} = 0.0959$ kJ/mol, $\Delta S_{\text{ex}} = 200.668$ J/mol and $\Delta G_{\text{ex}} = -77.655$ kJ/mol.

Then, under selected conditions for all chemical concentrations and temperatures, the CPE procedure was used to determine various heating times ranging from 5 to 30 minutes (Figure 6). The selected heating time was 10 minutes. Heating time offers the required energy to receive the right rate of thermodynamic equilibrium for the synthesis of solvation species, and higher extraction efficiency [14,15].

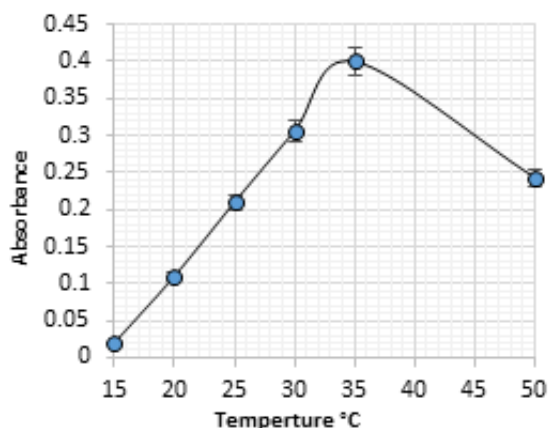


Figure 5. Influence of the temperature on extraction efficiency.

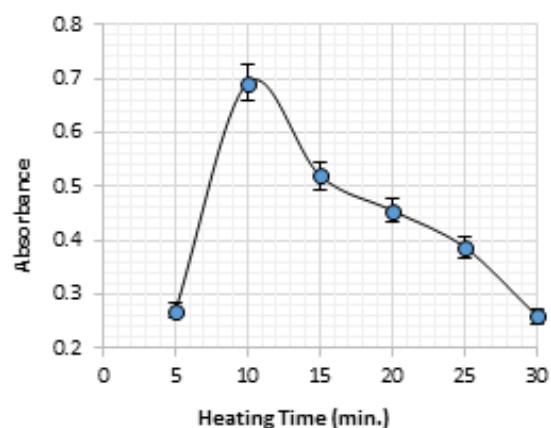


Figure 6. Influence of the heating time on extraction efficiency.

Influence of the Triton X-114 volume

Some features of the non-ionic surfactant Triton X-114, such as lipophilicity and ability to endure the electrolytes caused by corresponding silver ions, enabled the CPE and analytical selectivity for silver ions [33,34]. Figures 7 and 8 show the influence of Triton X-114 (1.0% v/v) at various volumes (0.1-1.0 mL). Better results, such as a high extraction efficiency, were obtained with 0.5 mL of Triton X-114, as the critical micellar concentration (CMC) was surpassed.

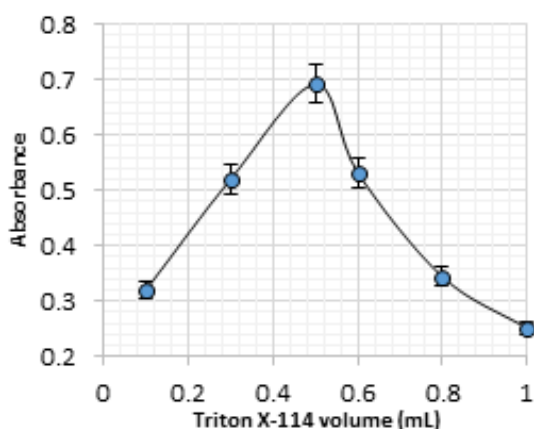


Figure 7. Influence of the Triton X-114 volume on the absorbance of extracted species.

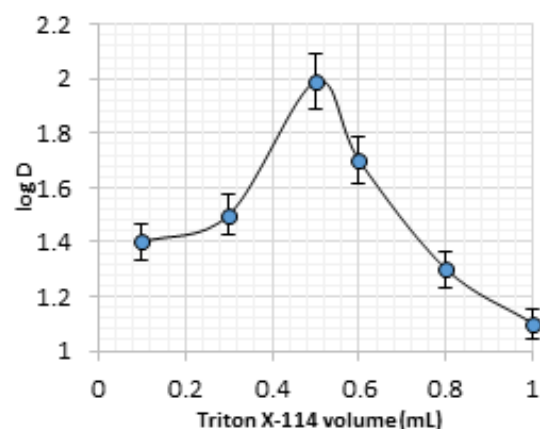


Figure 8. Influence of the Triton X-114 volume effect on D.

Comparison with other procedures

Some parameters of the proposed CPE, such as organic ligands, LOD and E were compared with those related to other strategies for preconcentration of silver ions using Triton X-114 as a surfactant [25, 33-44]. Table II summarizes the results. It can be seen that the LOD of the present method is lower, in relation to other procedures [35,37,39,45], whereas the best LOD was obtained by the CPE method [36]. The large and excellent E is probably due to the higher extraction efficiency of silver ions.

Table II. Comparison of Procedures for Silver Determination using CPE and Triton X-114

Organic ligands	LOD	E	References
2,4-DMP	0.05 $\mu\text{g L}^{-1}$	200	this work
6-(4-BrPAA)	0.0054 $\mu\text{g mL}^{-1}$	12.4	[44]
AgNPs	1.50 ng L^{-1}	-	[38]
APDC	0.42 $\mu\text{g L}^{-1}$	20	[35]
BIES	1.40 ng mL^{-1}	42	[43]
BIMPI	10.00 $\mu\text{g L}^{-1}$	35	[37]
BMAA	0.43 ng mL^{-1}	-	[34]
DDTC	1.00 $\mu\text{g L}^{-1}$	24	[45]
DDTC	0.30 ng mL^{-1}	33	[36]
H ₂ Dz	0.56 ng mL^{-1}	43	[40]
HPCTS	0.08 $\mu\text{g L}^{-1}$	46	[25]
MBT	2.20 ng mL^{-1}	-	[33]
PAR	6.00 $\mu\text{g L}^{-1}$	-	[39]

6-(4-BrPAA) 6-(4-bromo-phenylazo)m-anisidine, AgNPs silver nanoparticles, APDC ammonium pyrrolidine dithiocarbamate, BIES bis((1H-benzo [d] imidazol-2yl)ethyl) sulfane, BIMPI 2-((2-((1H-benzo[d]imidazole-2-yl)methoxy)phenoxy)methyl)-1H-benzo[d]imidazol, BMAA bis(2-mercaptoanil) acetylacetone, DDTC diethyl dithiocarbamate, H₂Dz Dithiazone, HPCTS 4-(p-chlorophenyl)-1-(pyridin-2-yl)thiosemicarbazide, MBT 2-mercaptobenzothiazole, PAR 4,2 pyridylazo resorsinol.

Analytical Figures of Merit

Limit of detection, limit of quantification, enrichment factor, precision, and linearity of the analytical curve were also examined, and the results are summarized in Table III. Analysis of this table reveals that silver recoveries were quantitative (> 98.7%), so the analytical accuracy was good. For $C = 30 \mu\text{g L}^{-1}$, RSD was 1.5%, based on five-fold assaying times of a 50 mL of solution. Under ideal conditions, the linearity was notes within 0.1 and $100 \mu\text{g L}^{-1}$. The standard error of detection limit detection limit for the blank solution (3.3 S/b) was determined as $0.05 \mu\text{g L}^{-1}$. Instead, the E was estimated as 200, by using the ratio of the slopes of the analytical curve after and before CPE.

Table III. Analytical Figures of Merit for CPE

Parameters	CPE
Enrichment factor	200
Limit of Detection	$0.050 \mu\text{g L}^{-1}$
Limit of Quantification	$0.15 \mu\text{g L}^{-1}$
Linearity	$0.1-100 \mu\text{g L}^{-1}$
Recovery	99.2-107.1%
Relative Standard Deviation (n = 5)	0.2-3.9%

CPE conditions: 50 mL sample, 1% (v/v) Triton X-114 (0.5 mL), $0.4 \text{ mol L}^{-1} \text{ NaNO}_3$, $1 \times 10^{-4} \text{ mol L}^{-1}$ 2,4-DMP, temperature $35 \text{ }^\circ\text{C}$ and time 10 min.

Applications

The proposed innovation was applied to silver determination in natural waters and sand tests from Al-Najaf city. Accuracy was assessed by analyzing samples already run by other methods. CPE was used to allow the estimation of recoveries after addition of known amounts of silver ions to the samples. The results (Table IV) demonstrate the CPE ability to extract and determine silver ions in water tests with various materials in the appropriate conditions, as well its ability to isolate silver ions. Results of silver of determination by two classical methods (FAAS and UV-Vis spectrophotometry) are also presented. Table V summarizes the obtained results.

Table IV. Silver determination in river and rain water samples three times

Tests	Added ($\mu\text{g L}^{-1}$)	CPE			FAAS ($\mu\text{g L}^{-1}$)	Dithizone ($\mu\text{g L}^{-1}$)
		Found ($\mu\text{g L}^{-1}$)	RSD (%)	Recovery (%)		
	-	N.D.	-	-		
Shatt Al-Kufa water	2.00	1.63	2.10	99.22	2.10	1.90
	4.00	4.11	0.23	107.10	4.02	3.85
	6.00	5.88	1.47	99.30	6.01	6.10
	-	N.D.	-	-		
Rainwater ^b	2.00	2.14	0.12	98.50	2.00	1.78
	4.00	4.40	1.23	102.50	4.80	4.82
	6.00	6.06	1.50	100.60	6.39	6.31

^bCollected at Al-Najaf city, Iraq (December 2020). N.D. not detected.

Table V. Silver determination in sand samples by the proposed innovation

Samples*	mean \pm RSD (n = 5) $\mu\text{g L}^{-1}$ Ag
1	3.02 \pm 3.40
2	5.88 \pm 1.60
3	3.38 \pm 1.40
4	4.25 \pm 2.90
5	5.01 \pm 0.70
6	3.33 \pm 2.00

*These sand samples were collected from various locations in the Al-Najaf desert.

CONCLUSIONS

An environmentally friendly, low-cost, and selective procedure exploiting CPE was proposed for separating and determining silver ions in water and sand samples. All reagents and solvents were safe for the environment. Outstanding limits of detection and quantification, and analytical recovery were attained. The procedure involves a short separation and preconcentration time (10 minutes). For the analysis of several samples with different concentrations, the advantages of combining CPE (easy, safe, rapid, and efficient) and the solvation process utilizing a lipophilic agent (selectivity and sensitivity) for silver ions were demonstrated. For the fast extraction of silver in biological samples, the developed strategy produced excellent results in terms of selectivity, environmental safety, simplicity, and accuracy.

Conflicts of interest

The authors declare no conflict of interest.

Acknowledgements

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